PCRBIOSYSTEMS
simplifying research

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qPCRBIO SyGreen® Mix Lo-ROX

Product description

Combined with the latest advancements in polymerase technology and advanced buffer chemistry, qPCRBIO SyGreen® Mix offers market-leading performance with minimal optimisation. The mix uses a proprietary intercalating dye that does not inhibit PCR, unlike other popular dyes.

qPCRBIO SyGreen® Mix uses antibody-mediated hot start technology that prevents the formation of primer-dimers to improve reaction sensitivity and specificity.

High-throughput screening has resulted in a buffer system that allows efficient amplification from GC-rich and AT-rich templates, under fast and standard cycling conditions.

Quality control

PCR Biosystems operates under an ISO 13485 certified Quality Management System. Our products are extensively tested and undergo a comprehensive, multi-step quality control process according to ISO 13485 standards, to ensure optimum performance, consistency and traceability.

Pack size	2x qPCRBIO SyGreen® Mix Lo-ROX
100 reactions	1 x 1 mL
500 reactions	5 x 1 mL
2000 reactions	20 x 1 mL
5000 reactions	1 x 50 mL bottle
5000 reactions	50 x 1 mL tubes

Shipping and storage

On arrival the kit should be stored between -30 °C and -20 °C. Avoid prolonged exposure to light. If stored correctly, the kit will retain full activity until the indicated expiry date. The kit can be stored at 4 °C for 1 month.

Technical support

Scan or click the QR code for troubleshooting help and answers to frequently asked technical questions. For further technical support, please email technical@pcrbio.com with the following information:

- Amplicon size
- Reaction setup
- Cycling conditions
- Screen grabs of amplification traces and melting profile

TROUBLESHOOT

FAQS

Product Use: Unless we agree otherwise in writing, the Goods we supply are provided:

1. For research purposes only and you should not use or rely on the Goods for diagnostic purposes. If you wish to use the Goods in a regulatory approved medical device, please contact us so that we may consider this and discuss it further with you.
2. Subject to our standard terms and conditions found at https://pcrbio.com/terms-conditions/.

Important considerations

Instrument compatibility: Different qPCR instruments may require different levels of ROX passive reference for normalisation. Use our qPCR Selection Tool to determine which ROX concentration your instrument requires (https://pcrbio.com/resources/qpcr-selection-tool/).

Primer design: For efficient amplification we recommend amplicon lengths between 80-200 bp and not exceeding 400 bp. Shorter amplicons allow for faster cycling. Primers should have an approximate Tm of ~60 ˚C using default Primer 3 settings (https://bioinfo.ut.ee/primer3/). To verify the best annealing temperature for your primers in our products, please visit https://pcrbio.com/resources/tm-calculator/.

Template amount: The kit can be used with cDNA or DNA synthesised or extracted by most commercial kits or standard extraction methods, provided the amount and quality of template are within an acceptable range. Addition of 2 to 5 µL volumes of sample will improve assay precision. For genomic DNA, 1 µg or less is recommended. For cDNA, 100 ng or less is recommended. However, users are encouraged to attempt a dilution series for new template/primer pairs to ensure that the PCR is efficient at that template concentration.

Reaction setup

1. Before starting, briefly vortex 2x qPCRBIO SyGreen® Mix.
2. Prepare a master mix based on following table:

Reagent	20 µL reaction	Final concentration	Notes
2x qPCRBIO SyGreen® Mix	10 µL	1x	
Forward primer (10 µM)	0.8 µL	400 nM	See above for optimal primer design
Reverse primer (10 µM)	0.8 µL	400 nM	
Template DNA	<100 ng cDNA, <1 µg genomic	variable	See above for template considerations
PCR grade dH_2O	Up to 20 µL final volume		

3. Program the instrument using following conditions, acquiring data on the FAM channel:

Cycles	Temperature	Time	Notes
1	95 ˚C	2 min	Polymerase activation, 2 minutes for cDNA and 3 minutes for genomic
40	95 ˚C 60 ˚C to 65 ˚C	5 seconds 20-30 seconds	Denaturation Anneal/Extension, do not exceed 30 seconds, do not use temperatures below 60 ˚C
Melt analysis	Refer to instrument instructions		Optional melt profile analysis