Directions for Use

RNA EZ-Vision® Loading Buffer, 1.5X

Code	Description	Size
N717-2X1.5ML	**RNA EZ-Vision® Loading Buffer, 1.5X**	2 x 1.5 mL tubes
N717-Q-SAMPLE	**RNA EZ-Vision® Loading Buffer, 1.5X**	1 x 0.3 mL tube
N717-SAMPLE	**RNA EZ-Vision®, RNA Dye As Loading Buffer (1.5X)**	1 x 0.3 mL tube

General Information

VWR Life Science AMRESCO's RNA EZ-Vision® is a convenient, easy-to-use alternative to ethidium bromide for immediate visualization of RNA bands in denaturing formaldehyde agarose gels. The non-mutagenic, fluorescent stain, supplied in a loading buffer, binds to the RNA sample prior to loading and co-migrates with it during electrophoresis. Brilliant green bands against a dark background are immediately visible upon illumination with a standard U.V. transilluminator, without the need for post-run staining and destaining. RNA EZ-Vision® is sensitive to approximately 150 ng RNA per band. Image capture can be performed with standard green filters.

The 1.5X loading buffer includes formamide as an RNA denaturant and bromophenol blue as a tracking dye. It is guaranteed free of RNase activity and is compatible with downstream applications including Northern blotting. RNA EZ-Vision® is ideal for environments needing to reduce ethidium bromide use. In addition it streamlines the number of steps required for RNA electrophoresis protocols.

Storage/Stability

Store at room temperature (18 to 26°C).

Product Use Limitations

For research use only. Not for therapeutic or diagnostic use.

Protocol/Procedure

Note: RNA EZ-Vision® is best used with RNA at a concentration of ≥ 1 mg/mL. If the stock RNA concentration is < 1.5 mg/mL, RNA EZ-Vision® should be used as a 2X solution instead of a 1.5X solution.

1. Vortex RNA EZ-Vision® Loading Buffer prior to use to ensure the solution is homogenous.
2. Add 1 volume of RNA sample to 2 volumes of RNA EZ-Vision® Loading Buffer (e.g. 5 µL of RNA sample mixed with 10 µL of Vortex RNA EZ-Vision® Loading Buffer).
3. Heat denature the samples for 10 minutes at 65°C.
4. Load the heat denatured RNA samples into a 1 – 2% formaldehyde denaturing gel.
5. Separate the RNA at 5 – 8 V/cm according to standard protocols.
6. After the run, remove gel and place on UV transilluminator to immediately visualize the RNA.
7. For optimal results, image capture should be conducted with a SYBR® Green filter (green emission filter).

For Technical Support

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